

Review Article

Landscape of Genetic Alterations in Patients with Acute Myeloid Leukemia and the Implications in Personalized Medicine

Hwei-Fang Tien^{1,2}

¹Department of Internal Medicine, National Taiwan University, Taipei, Taiwan

²Department of Internal Medicine, Far Eastern Memorial Hospital, New Taipei, Taiwan

Abstract

Genetic alterations play important roles in the pathogenesis of acute myeloid leukemia (AML). With advances in genomics, many gene mutations have been detected in AML, and some specific mutations are included in international classifications and risk stratification. Gene mutations can be targets for the development of novel agents and biomarkers for monitoring measurable/minimal residual disease (MRD). Personalized medicine according to the genetic risk at presentation and MRD after treatment can not only improve survival but also reduce toxicity from therapies. This review focuses on the landscape of gene mutations and their clinical implications in patients with AML.

Keywords: Acute myeloid leukemia, gene mutation, measurable/minimal residual disease

INTRODUCTION

Acute myeloid leukemia (AML) is a heterogeneous group of neoplastic diseases with great variability in the pathogenesis, clinical presentations, and treatment responses.^[1] It is characterized by the uncontrolled proliferation of hematopoietic progenitors and the inability of cells to differentiate. Patients usually present with anemia, infection, and bleeding. About 60%–70% of patients can achieve complete remission (CR) after intensive chemotherapy (IC), but most relapse within 1 year, indicating that there is still minimal/measurable residual disease (MRD)

after chemotherapy^[2] even though the patients are in morphological CR.

Personalized treatment according to the AML pathogenesis and risk factors of individual patients is needed to improve the survival of AML patients while reducing the side effects from treatment. The risk factors include those that are related to the patient such as age and factors related to the disease such as white blood cell count, cytogenetics,

Address for correspondence: Prof. Hwei-Fang Tien,
Department of Internal Medicine, National Taiwan University, No. 7
Chung-Shan South Road, Taipei, Taiwan.
E-mail: hftien@ntu.edu.tw

Submitted: 06-Jan-2025

Revised: 28-Feb-2025

Accepted: 27-Mar-2025

Published: 27-Jun-2025

Access this article online

Quick Response Code:



Website:
<https://journals.lww.com/jcrp>

DOI:
10.4103/ejcrp.eJCRP-D-25-00001

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: WKHLRPMedknow_reprints@wolterskluwer.com

How to cite this article: Tien HF. Landscape of genetic alterations in patients with acute myeloid leukemia and the implications in personalized medicine. *J Cancer Res Pract* 2025;12:33-9.

and molecular gene mutations.^[3-5] This review focuses on molecular gene mutations and their implications in personalized treatment.

LANDSCAPE OF GENETIC ALTERATIONS IN ACUTE MYELOID LEUKEMIA

Recurrent cytogenetic abnormalities, such as t (15;17)/*PML::RARA*, t (8;21)/*RUNX1::RUNX1T1*, inv (16)/*CBFB::MYH11*, and del/t (11)(q23)/*KMT2A* fusion, have long been known to play important roles in the pathogenesis of AML and are closely associated with clinical outcomes.^[3] With advances in genomic techniques, a variety of molecular mutations which cannot be detected by conventional chromosomal analysis have been found. Some of these mutations have been shown to contribute to the development and progression of AML and predict clinical outcomes.^[4]

More than 90% of AML patients have molecular aberrations, including mutations in *ASXL1*, *CEBPA*, *DNMT3A*, *EZH2*, *FLT3*-internal tandem duplication (ITD), *IDH1/IDH2*, *NPM1*, *PTPN11*, *RUNX1*, *WT1*, *TET2*, *TP53*, RNA splicing factor genes (such as *SF3B1*, *SRSF2*, and *U2AF1*), cohesin complex genes (such as *STAG1*, *STAG2*, and *RAD21*), etc.^[6-17] Table 1 summarizes the common gene mutations in AML according to functional categories, and Figure 1 shows the distribution of gene mutations in newly diagnosed AML patients from the National Taiwan University Hospital (NTUH). The two most common mutations are *FLT3*-ITD and *NPM1* mutation, followed by *DNMT3A* mutation. It is common to observe two or more mutations occurring in the same patient, consistent with the two-hit theory. Some mutations frequently co-occur, such as *NPM1* mutation, *FLT3*-ITD, and *DNMT3A* mutation, indicating their concert interaction in the pathogenesis of AML. Conversely, others such as *IDH* and *TET2* mutations

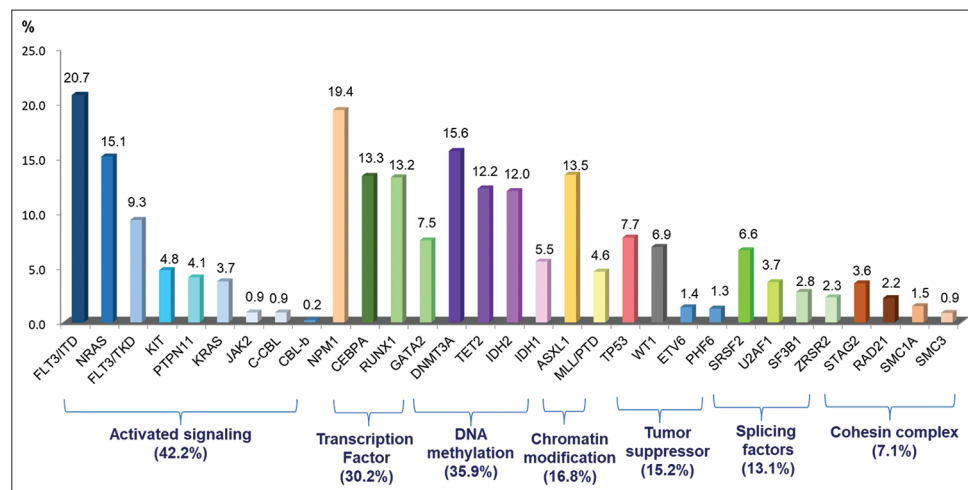


Figure 1: Common molecular gene mutations and their incidence rates (from the acute myeloid leukemia cohort at National Taiwan University Hospital, $n = 763$). ITD: Internal tandem duplication

Table 1: Common genetic alterations in acute myeloid leukemia according to functional categories

Functional category	Gene members	Role in AML leukemogenesis
Myeloid transcription factors	Transcription factor fusions by chromosomal rearrangements, such as t(8;21)(q22;q22)/ <i>RUNX1::RUNX1T1</i> and inv(16)(p13.1;q22) or t(16;16)(p13.1;q22)/ <i>CBFB::MYH11</i> <i>GATA2</i> , <i>RUNX1</i> , and <i>CEBPA</i>	Transcriptional deregulation and impaired hematopoietic differentiation
<i>NPM1</i>	<i>NPM1</i>	Aberrant cytoplasmic localization of <i>NPM1</i> and its interacting proteins
Tumor suppressors	<i>TP53</i> , <i>WT1</i> , <i>PHF6</i>	Transcriptional deregulation and impaired degradation via the negative regulator (MDM2 and PTEN oncogenes)
Signaling pathways	<i>FLT3</i> , <i>KIT</i> , <i>PTPN11</i> , <i>RAS</i>	Proliferative advantage through the RAS-RAF, JAK-STAT, and PI3K-AKT signaling pathways
DNA methylation	<i>DNMT3A</i> , <i>TET2</i> , <i>IDH1</i> , <i>IDH2</i>	Deregulation of DNA methylation and oncometabolite production
Chromatin modifier	<i>ASXL1</i> , <i>EZH2</i> and <i>KMT2A</i>	Deregulation of chromatin modification and impairment of methyltransferase function
Cohesin complex	<i>STAG1</i> , <i>STAG2</i> , <i>RAD21</i> , <i>SMC1A</i> , <i>SMC3</i>	Impairment of accurate chromosome segregation and transcriptional regulation
Splicing factors	<i>SRSF2</i> , <i>SF3B1</i> , <i>U2AF1</i> , <i>ZRSR2</i>	Deregulated RNA processing and aberrant splicing patterns

NPM1: Nucleophosmin, AML: Acute myeloid leukemia

are mutually exclusive, indicating their redundant action in AML cells.

INCORPORATION OF MOLECULAR GENE MUTATIONS INTO THE CLASSIFICATION OF ACUTE MYELOID LEUKEMIA

In the 2001 World Health Organization (WHO) classification of myeloid neoplasms, only cytogenetic abnormalities were included as recurrent genetic abnormalities in AML.^[18] With molecular mutations detected in the following years, the 2008 WHO classification first included AML with two molecular mutations, *NPM1* mutation and biallelic *CEBPA* mutation, respectively, as provisional entities.^[19] The list of AML entities with molecular mutations increased in the 2016 WHO classification^[20] and expanded further in the latest 2022 WHO classification^[21] [Table 2]. There are some differences between the 2022 International Consensus Classification (ICC) of myeloid neoplasms and the 2022 WHO classification^[22] [Table 3]. The major difference is the reclassification of myelodysplastic syndrome (MDS)

with 10%–19% blasts in the blood or bone marrow to MDS/AML. The purpose of this change is to allow MDS/AML patients to be eligible for both MDS and AML trials to optimize their management. AML with specific recurrent gene fusions or *NPM1* mutation shows distinct clinical and biological characteristics irrespective of blast counts in the bone marrow or peripheral blood (BM or PB), so the cutoff level of blast count for the diagnosis of AML was reduced from $\geq 20\%$ in the 2016 WHO classification to $\geq 10\%$ in the 2022 ICC [Table 3], and there is no lower limit in the 2022 WHO classification [Table 2] in these AML categories, with the exception of AML with *BCR::ABL* fusion in which a blast cutoff of 20% is required to avoid overlap with chronic myeloid leukemia [Tables 2 and 3]. The same blast cutoff of 10% is applied for AML with *CEBPA* mutation in the ICC classification; however, the cutoff is 20% in the WHO classification, because the WHO panelists considered that there are insufficient data to support any change in the blast cutoff criterion for AML with *CEBPA* mutation.

RISK CLASSIFICATION BASED ON GENE MUTATIONS

There are close associations of specific molecular mutations with the clinical and biological features of AML. For example,

Table 2: The 2022 World Health Organization Classification

AML with defining genetic abnormalities
APL with <i>PML::RARA</i> fusion*
AML with <i>RUNX1::RUNX1T1</i> fusion*
AML with <i>CBFB::MYH11</i> fusion*
AML with <i>DEK::NUP214</i> fusion*
AML with <i>RBM15::MRTF</i> A fusion*
AML with <i>BCR::ABL1</i> fusion
AML with <i>KMT2A</i> rearrangement*
AML with <i>MECOM</i> rearrangement*
AML with <i>NUP98</i> rearrangement*
AML with <i>NPM1</i> mutation*
AML with <i>CEBPA</i> mutation
AML, myelodysplasia-related**
AML with other defined genetic alterations
AML, defined by differentiation
AML with minimal differentiation
AML without maturation
AML with maturation
Acute basophilic leukemia
Acute myelomonocytic leukemia
Acute monocytic leukemia
Acute erythroid leukemia
Acute megakaryoblastic leukemia

*Blasts $<20\%$ in the marrow or PB is acceptable for the diagnosis of AML with defining genetic abnormalities except AML with *BCR::ABL1* fusion and AML with *CEBPA* mutation, **Defining cytogenetic abnormalities: Complex karyotype (≥ 3 abnormalities); 5q deletion or loss of 5q due to unbalanced translocation; Monosomy 7, 7q deletion, or loss of 7q due to unbalanced translocation; 11q deletion; 12p deletion or loss of 12p due to unbalanced translocation; Monosomy 13 or 13q deletion; 17p deletion or loss of 17p due to unbalanced translocation; Isochromosome 17q; idic(X)(q13). Defining somatic mutations: *ASXL1*, *BCOR*, *EZH2*, *RUNX1*, *SF3B1*, *SRSF2*, *STAG2*, *U2AF1*, or *ZRSR2*. Khoury *et al.*, 2022.^[21] AML: Acute myeloid leukemia, APL: Acute myeloid leukemia

Table 3: The 2022 International Consensus Classification

APL with t(15;17)(q24.1;q21.2)/ <i>PML::RARA</i> $\geq 10\%*$
APL with other <i>RARA</i> rearrangements $\geq 10\%*$
AML with t(8;21)(q22;q22.1)/ <i>RUNX1::RUNX1T1</i> $\geq 10\%*$
AML with inv(16)(p13.1;q22) or t(16;16)(p13.1;q22)/ <i>CBFB::MYH11</i> $\geq 10\%*$
AML with t(9;11)(p21.3;q23.3)/ <i>MLLT3::KMT2A</i> $\geq 10\%*$
AML with other <i>KMT2A</i> rearrangements $\geq 10\%*$
AML with t(6;9)(p22.3;q34.1)/ <i>DEK::NUP214</i> $\geq 10\%*$
AML with inv(3)(q21.3;q26.2) or t(3;3)(q21.3;q26.2)/ <i>GATA2;MECOM(EV1)</i> $\geq 10\%*$
AML with other <i>MECOM</i> rearrangements $\geq 10\%*$
AML with other rare recurring translocations $\geq 10\%*$
AML with t(9;22)(q34.1;q11.2)/ <i>BCR::ABL1</i>
AML with mutated <i>NPM1</i> $\geq 10\%*$
AML with in-frame bZIP <i>CEBPA</i> mutations $\geq 10\%*$
AML and MDS/AML with mutated <i>TP53</i>
AML and MDS/AML with myelodysplasia-related gene mutations**
AML with myelodysplasia-related cytogenetic abnormalities#
AML and MDS/AML NOS
Myeloid sarcoma

*The diagnosis of AML with these recurrent genetic alterations can be made when blasts (or so-called “blast equivalents” including promonocytes and neoplastic promyelocytes) account for 10% or more in the blood or marrow, **Defined by mutations in *ASXL1*, *BCOR*, *EZH2*, *RUNX1*, *SF3B1*, *SRSF2*, *STAG2*, *U2AF1*, or *ZRSR2*, #Defined by detecting a complex karyotype (≥ 3 unrelated clonal chromosomal abnormalities), del(5q)/t(5q)/add(5q), -7/del(7q), 18, del(12p)/t(12p)/add(12p), i(17q), -17/add(17p) or del(17p), del(20q), and/or idic(X)(q13) clonal abnormalities. Arber *et al.*, 2022.^[22] APL: Acute promyelocytic leukemia, AML: Acute myeloid leukemia, *NPM1*: Nucleophosmin, MDS: Myelodysplastic syndrome, NOS: Not otherwise specified, bZIP: Basic leucine zipper

CEBPA in-frame basic leucine zipper domain mutations and *NPM1* mutation in the absence of *FLT3*-ITD predict longer survival; in contrast, *ASXL1*, *BCOR*, *EZH2*, *RUNX1*, *SF3B1*, *SRSF2*, *STAG2*, *U2AF1*, *ZRSR2* mutations (all belonging to myelodysplasia-related gene mutations), and *TP53* mutations predict poorer outcomes.^[7,23] After the publication of the 2022 WHO and ICC classifications, European LeukemiaNet (ELN) updated the prior 2017^[24] ELN recommendations for the diagnosis and management of AML in 2022.^[25] In the recommendations, AML is risk stratified according to cytogenetic abnormalities and gene mutations into three categories: favorable, intermediate, and adverse [Table 4]. Patients with different risks have distinct clinical outcomes. Figure 2 shows overall survival curves stratified by the 2022 ELN risk classification of 809 AML patients from NTUH.^[26]

GENE MUTATIONS AS TARGETS FOR THE DEVELOPMENT OF NOVEL AGENTS

Gene mutations can be targets for novel therapies, and several agents targeted to specific genetic alterations have been developed.^[27-35] Some are available in Taiwan, including

Table 4: 2022 European LeukemiaNet risk classification by genetics

Risk category	Genetic abnormality
Favorable	t(8;21)(q22;q22.1)/ <i>RUNX1::RUNX1T1</i> inv(16)(p13.1;q22) or t(16;16)(p13.1;q22)/ <i>CBFB::MYH11</i> Mutated <i>NPM1</i> without <i>FLT3</i> -ITD bZIP in-frame mutated <i>CEBPA</i>
Intermediate	Mutated <i>NPM1</i> with <i>FLT3</i> -ITD Wild-type <i>NPM1</i> with <i>FLT3</i> -ITD t(9;11)(p21.3;q23.3)/ <i>MLL2::KMT2A</i> Cytogenetic and/or molecular abnormalities not classified as favorable or adverse
Adverse	t(6;9)(p23;q34.1)/ <i>DEK::NUP214</i> t(v;11q23.3)/ <i>KMT2A</i> -rearranged t(9;22)(q34.1;q11.2)/ <i>BCR::ABL1</i> t(8;16)(p11;p13)/ <i>KAT6A::CREBBP</i> inv(3)(q21.3;q26.2) or t(3;3)(q21.3;q26.2)/ <i>GATA2, MECOM(EV11)</i> t(3q26.2;v)/ <i>MECOM(EV11)</i> -rearranged* -5 or del(5q); -7; -17/abn(17p) Complex karyotype, monosomal karyotype [#] Mutated <i>ASXL1</i> , <i>BCOR</i> , <i>EZH2</i> , <i>RUNX1</i> , <i>SF3B1</i> , <i>SRSF2</i> , <i>STAG2</i> , <i>U2AF1</i> , or <i>ZRSR2</i> Mutated <i>TP53</i>

*t(3q26.2;v)/*MECOM(EV11)*-rearranged: 3q26.2 fusions with partners than *GATA2*, such as t(3;8)(q26.2;q24.2)/*MYC::MECOM*; t(3;12)(q26.2;p13.2)/*ETV6::MECOM*; t(3;21)(q26.2;q22.1)/*MECOM::RUNX1MYC*, etc., are also associated with poor prognosis. [#]Monosomal karyotype: Presence of two or more distinct monosomies (excluding loss of X or Y), or one single autosomal monosomy in combination with at least one structural chromosome abnormality (excluding core-binding factor AML). Döhner *et al.*, 2022.
[25] AML: Acute myeloid leukemia, *NPM1*: Nucleophosmin, ITD: Internal tandem duplication, bZIP: Basic leucine zipper

the *FLT3* inhibitors midostaurin and gilteritinib^[28,29] for *FLT3*-mutated AML, while some others have been approved by the US Food and Drug Administration (FDA) but not yet by the Taiwan FDA, such as the *IDH1*/*IDH2* inhibitors, ivosidenib and enasidenib, respectively, for *IDH*-mutated AML.^[33,35] In November 2024, the US FDA approved the first menin inhibitor, revumenib, for relapsed or refractory acute leukemia with *KMT2A* translocation (<https://www.fda.gov/drugs/resources-information-approved-drugs/fda-approves-revumenib-relapsed-or-refractory-acute-leukemia-kmt2a-translocation>). More novel agents are under clinical trials, including other menin inhibitors for *KMT2A*-rearranged or *NPM1*-mutated AML, eprenetapopt, a reactivator of p53, for *TP53*-mutated AML,^[30] and H3B-8800, an orally active modulator of SF3B1 splicing complex, for spliceosome-mutant cancers.^[36]

GENE MUTATIONS AS BIOMARKERS FOR MONITORING MEASURABLE/MINIMAL RESIDUAL DISEASE

Most AML patients can achieve cytomorphological CR; however, approximately 50% eventually relapse, indicating the presence of MRD that escapes the detection of conventional morphological examinations. Mounting evidence has shown that a persistently high level of MRD or a rising level after an initial response invariably predicts relapse.

Traditionally, MRD is detected by multicolor flow cytometry, which recognizes leukemia-associated or a different from normal aberrant immunophenotype as a leukemia-specific marker.^[37] Quantifying MRD by reverse quantitative polymerase chain reaction is based on AML-specific fusion genes such as *PML::RARA*, *RUNX1::RUNX1T1*, and *CBFB::MYH11* or gene mutations such as *NPM1* mutation. Next-generation sequencing is a more powerful technique which can simultaneously detect various mutations and be applied to most AML patients.^[38]

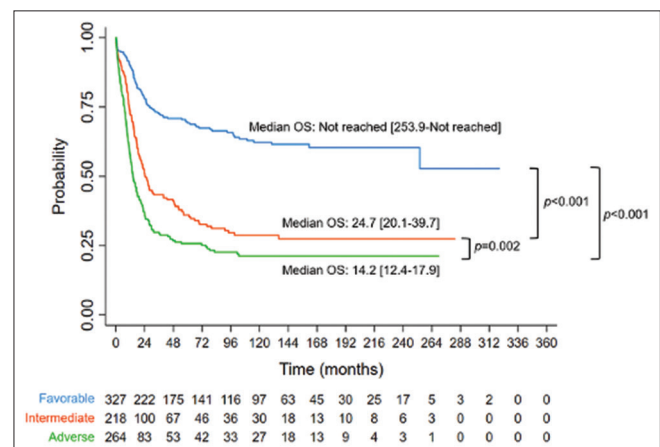


Figure 2: Overall survival of acute myeloid leukemia (AML) patients stratified by the European LeukemiaNet 2022 risk classification (from the AML cohort at National Taiwan University Hospital, $n = 809$).^[26] OS: Overall survival

IMPLICATIONS OF GENE MUTATIONS ON PERSONALIZED TREATMENT

Identifying the prognostic factors for patients with AML is the first step of personalized medicine. Conventionally, AML patients are categorized into favorable-, intermediate-, and unfavorable-cytogenetic risk groups according to cytogenetic changes [Figure 3]. However, patients with intermediate-risk cytogenetics represent a largely heterogeneous population regarding treatment response and clinical outcome. Integrating gene mutations with cytogenetic abnormalities can better stratify AML patients into different risk groups as recommended by the ELN.^[25] Patients with favorable-risk AML can be

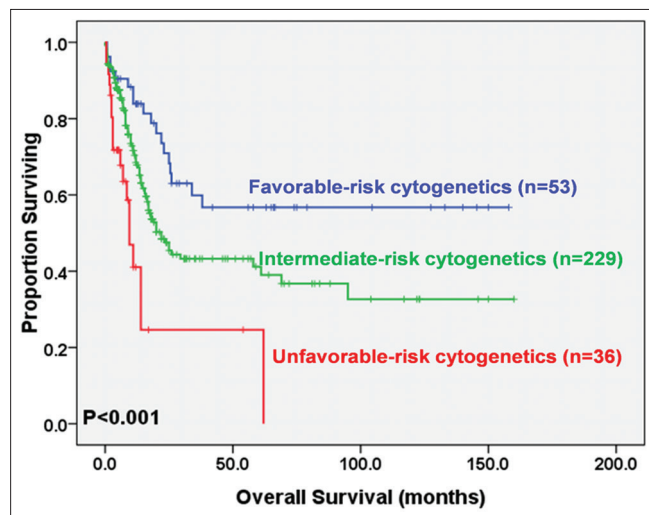


Figure 3: Overall survival of acute myeloid leukemia patients from National Taiwan University Hospital stratified by cytogenetic risk. Favorable-risk cytogenetics: t (8;21), inv (16), t (15;17); unfavorable-risk cytogenetics: Complex, -5/5q-, -7/7q-, 3q-, t (9;22)/(Ph); intermediate-risk cytogenetics: Normal karyotype or all other abnormalities^[39]

treated with consolidation chemotherapy after achieving CR with no need to proceed to allogeneic hematopoietic transplantation (allo-HSCT). However, the treatment strategy should be changed if MRD is detected during follow-up, even if the patient has good-risk genetic aberrations. On the other side, patients with adverse-risk AML can benefit from IC followed by allo-HSCT. For example, AML patients with *RUNX1* mutation have poorer outcomes than those without the mutation; however, we previously showed that *RUNX1*-mutated patients had similar survival to *RUNX1*-wild patients if receiving allo-HSCT.^[40] Similarly, allo-HSCT has been shown to ameliorate the poor prognosis of AML patients with myelodysplasia-related mutations.^[23]

MRD after treatment is an important biomarker to monitor treatment response, detect early relapse, and predict the prognosis in AML patients. We previously showed that MRD positivity detected by NGS at first morphological CR after induction chemotherapy [Figure 4, left] as well as first consolidation chemotherapy [Figure 4, right] predicts poorer outcomes in AML patients,^[38] especially at the latter time point. The poor prognosis of patients with NGS MRD after consolidation chemotherapy can be mitigated by allo-HSCT. Preemptive treatment with azacitidine^[41] or venetoclax, an oral BCL-2 inhibitor,-based treatment^[42] when MRD is detected is effective to reduce or eliminate MRD.

The use of novel agents targeting specific gene mutations is expected to improve the treatment response and clinical outcomes of AML patients.^[43] *FLT3*-mutated AML patients have been shown to have longer survival if receiving frontline IC plus the FLT3 inhibitor midostaurin, compared to IC alone.^[28] Furthermore, the addition of ivosidenib, an IDH1 inhibitor, to the hypomethylation agent azacitidine has been shown to improve overall survival in newly diagnosed *IDH1*-mutated AML patients who are ineligible for IC compared

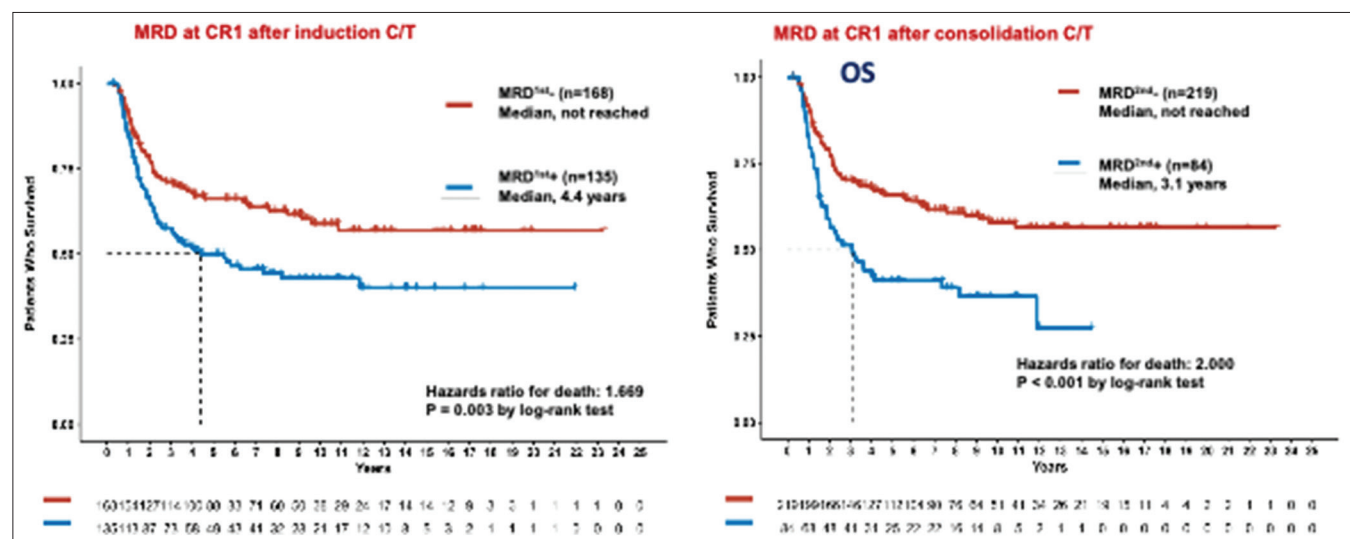


Figure 4: Overall survival curves stratified by the status of measurable/minimal residual disease (MRD) detected by next-generation sequencing at the time of morphological complete remission after induction chemotherapy (left) and first consolidation chemotherapy (right). MRD positivity predicts poorer overall survival, especially after first consolidation chemotherapy.^[38] OS: Overall survival

to treatment with azacitidine alone.^[44] Issa *et al.* reported that treatment with the menin inhibitor revumenib in patients with heavily pretreated relapsed/refractory acute leukemia with *KMT2A* translocation resulted in a rate of CR or CR with partial hematologic recovery of 22.8%.^[34] To achieve the goal of precision medicine, the development of more novel mutation-targeted agents is needed.

CONCLUSION

Genetic alterations are useful biomarkers to risk-stratify AML and guide treatment choice. They can also be targets for the development of novel agents and markers to monitor MRD. According to the 2022 ELN recommendation, in addition to searching for specific cytogenetic abnormalities/fusion genes such as *RUNX::RUNX1T1*, *CBFB::MYH11*, and *KMT2A* gene fusions, it is mandatory to screen for gene mutations that are important for AML classification and risk stratification, such as *NPM1*, *CEBPA*, *TP53*, *ASXL1*, *RUNX1*, *BCOR*, *EZH2*, *SF3B1*, *SRSF2*, *STAG2*, *U2AF1*, and *ZRSR2* mutations, and those that are druggable, such as *FLT3* and *IDH1/2* mutations. Personalized medicine based on the risk at diagnosis, MRD after treatment, and proper use of mutation-targeted novel agents can improve the survival of AML patients while reducing the toxicity from the treatment.

Data availability statement

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

- Döhner H, Weisdorf DJ, Bloomfield CD. Acute myeloid leukemia. *N Engl J Med* 2015;373:1136-52.
- Schuurhuis GJ, Heuser M, Freeman S, Béné MC, Buccisano F, Cloos J, *et al.* Minimal/measurable residual disease in AML: A consensus document from the European LeukemiaNet MRD working party. *Blood* 2018;131:1275-91.
- Grimwade D, Hills RK, Moorman AV, Walker H, Chatters S, Goldstone AH, *et al.* Refinement of cytogenetic classification in acute myeloid leukemia: Determination of prognostic significance of rare recurring chromosomal abnormalities among 5876 younger adult patients treated in the United Kingdom Medical Research Council trials. *Blood* 2010;116:354-65.
- Papaemmanuil E, Gerstung M, Bullinger L, Gaidzik VI, Paschka P, Roberts ND, *et al.* Genomic classification and prognosis in acute myeloid leukemia. *N Engl J Med* 2016;374:2209-21.
- Tien FM, Hou HA, Tsai CH, Tang JL, Chen CY, Kuo YY, *et al.* Hyperleukocytosis is associated with distinct genetic alterations and is an independent poor-risk factor in *de novo* acute myeloid leukemia patients. *Eur J Haematol* 2018;101:86-94.
- Chou WC, Chou SC, Liu CY, Chen CY, Hou HA, Kuo YY, *et al.* TET2 mutation is an unfavorable prognostic factor in acute myeloid leukemia patients with intermediate-risk cytogenetics. *Blood* 2011;118:3803-10.
- Hou HA, Chou WC, Kuo YY, Liu CY, Lin LI, Tseng MH, *et al.* TP53 mutations in *de novo* acute myeloid leukemia patients: Longitudinal follow-ups show the mutation is stable during disease evolution. *Blood Cancer J* 2015;5:e331.
- Chou WC, Hou HA, Chen CY, Tang JL, Yao M, Tsay W, *et al.* Distinct clinical and biologic characteristics in adult acute myeloid leukemia bearing the isocitrate dehydrogenase 1 mutation. *Blood* 2010;115:2749-54.
- Tsai CH, Hou HA, Tang JL, Kuo YY, Chiu YC, Lin CC, *et al.* Prognostic impacts and dynamic changes of cohesin complex gene mutations in *de novo* acute myeloid leukemia. *Blood Cancer J* 2017;7:663.
- Hou HA, Kuo YY, Liu CY, Chou WC, Lee MC, Chen CY, *et al.* DNMT3A mutations in acute myeloid leukemia: Stability during disease evolution and clinical implications. *Blood* 2012;119:559-68.
- Chou WC, Huang HH, Hou HA, Chen CY, Tang JL, Yao M, *et al.* Distinct clinical and biological features of *de novo* acute myeloid leukemia with additional sex comb-like 1 (*ASXL1*) mutations. *Blood* 2010;116:4086-94.
- Tang JL, Hou HA, Chen CY, Liu CY, Chou WC, Tseng MH, *et al.* AML1/RUNX1 mutations in 470 adult patients with *de novo* acute myeloid leukemia: Prognostic implication and interaction with other gene alterations. *Blood* 2009;114:5352-61.
- Hou HA, Huang TC, Lin LI, Liu CY, Chen CY, Chou WC, *et al.* WT1 mutation in 470 adult patients with acute myeloid leukemia: Stability during disease evolution and implication of its incorporation into a survival scoring system. *Blood* 2010;115:5222-31.
- Shiah HS, Kuo YY, Tang JL, Huang SY, Yao M, Tsay W, *et al.* Clinical and biological implications of partial tandem duplication of the MLL gene in acute myeloid leukemia without chromosomal abnormalities at 11q23. *Leukemia* 2002;16:196-202.
- Hou HA, Liu CY, Kuo YY, Chou WC, Tsai CH, Lin CC, *et al.* Splicing factor mutations predict poor prognosis in patients with *de novo* acute myeloid leukemia. *Oncotarget* 2016;7:9084-101.
- Chou WC, Tang JL, Lin LI, Yao M, Tsay W, Chen CY, *et al.* Nucleophosmin mutations in *de novo* acute myeloid leukemia: The age-dependent incidences and the stability during disease evolution. *Cancer Res* 2006;66:3310-6.
- Chou WC, Lei WC, Ko BS, Hou HA, Chen CY, Tang JL, *et al.* The prognostic impact and stability of isocitrate dehydrogenase 2 mutation in adult patients with acute myeloid leukemia. *Leukemia* 2011;25:246-53.
- Vardiman JW, Harris NL, Brunning RD. The World Health Organization (WHO) classification of the myeloid neoplasms. *Blood* 2002;100:2292-302.
- Vardiman JW, Thiele J, Arber DA, Brunning RD, Borowitz MJ, Porwit A, *et al.* The 2008 revision of the World Health Organization (WHO) classification of myeloid neoplasms and acute leukemia: Rationale and important changes. *Blood* 2009;114:937-51.
- Arber DA, Orazi A, Hasserjian R, Thiele J, Borowitz MJ, Le Beau MM, *et al.* The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. *Blood* 2016;127:2391-405.
- Khoury JD, Solary E, Abla O, Akkari Y, Alaggio R, Apperley JF, *et al.* The 5th edition of the World Health Organization classification of haematolymphoid tumours: Myeloid and histiocytic/dendritic neoplasms. *Leukemia* 2022;36:1703-19.
- Arber DA, Orazi A, Hasserjian RP, Borowitz MJ, Calvo KR, Kvasnicka HM, *et al.* International consensus classification of myeloid neoplasms and acute leukemias: Integrating morphologic, clinical, and genomic data. *Blood* 2022;140:1200-28.
- Tsai XC, Sun KJ, Lo MY, Tien FM, Kuo YY, Tseng MH, *et al.* Poor prognostic implications of myelodysplasia-related mutations in both older and younger patients with *de novo* AML. *Blood Cancer J* 2023;13:4.
- Döhner H, Estey E, Grimwade D, Amadori S, Appelbaum FR, Büchner T, *et al.* Diagnosis and management of AML in adults: 2017 ELN recommendations from an International expert panel. *Blood* 2017;129:424-47.
- Döhner H, Wei AH, Appelbaum FR, Craddock C, DiNardo CD, Dombret H, *et al.* Diagnosis and management of AML in adults: 2022 recommendations from an international expert panel on behalf of the ELN. *Blood* 2022;140:1345-77.
- Lo MY, Tsai XC, Lin CC, Tien FM, Kuo YY, Lee WH, *et al.* Validation

- of the prognostic significance of the 2022 European LeukemiaNet risk stratification system in intensive chemotherapy treated aged 18 to 65 years patients with *de novo* acute myeloid leukemia. *Am J Hematol* 2023;98:760-9.
27. Döhner H, Wei AH, Löwenberg B. Towards precision medicine for AML. *Nat Rev Clin Oncol* 2021;18:577-90.
 28. Stone RM, Mandrekar SJ, Sanford BL, Laumann K, Geyer S, Bloomfield CD, *et al.* Midostaurin plus chemotherapy for acute myeloid leukemia with a FLT3 mutation. *N Engl J Med* 2017;377:454-64.
 29. Perl AE, Martinelli G, Cortes JE, Neubauer A, Berman E, Paolini S, *et al.* Gilteritinib or chemotherapy for relapsed or refractory FLT3-mutated AML. *N Engl J Med* 2019;381:1728-40.
 30. Lambert JM, Gorzov P, Veprintsev DB, Söderqvist M, Segerbäck D, Bergman J, *et al.* PRIMA-1 reactivates mutant p53 by covalent binding to the core domain. *Cancer Cell* 2009;15:376-88.
 31. Krivtsov AV, Evans K, Gadrey JY, Eschle BK, Hatton C, Uckelmann HJ, *et al.* A Menin-MLL inhibitor induces specific chromatin changes and eradicates disease in models of MLL-rearranged leukemia. *Cancer Cell* 2019;36:660-73.e11.
 32. DiNardo CD, Stein EM, de Botton S, Roboz GJ, Altman JK, Mims AS, *et al.* Durable remissions with ivosidenib in IDH1-mutated relapsed or refractory AML. *N Engl J Med* 2018;378:2386-98.
 33. Roboz GJ, DiNardo CD, Stein EM, de Botton S, Mims AS, Prince GT, *et al.* Ivosidenib induces deep durable remissions in patients with newly diagnosed IDH1-mutant acute myeloid leukemia. *Blood* 2020;135:463-71.
 34. Issa GC, Aldoss I, Thirman MJ, DiPersio J, Arellano M, Blachly JS, *et al.* Menin inhibition with revumenib for KMT2A-rearranged relapsed or refractory acute leukemia (AUGMENT-101). *J Clin Oncol* 2025;43:75-84.
 35. Stein EM, DiNardo CD, Pollyea DA, Fathi AT, Roboz GJ, Altman JK, *et al.* Enasidenib in mutant IDH2 relapsed or refractory acute myeloid leukemia. *Blood* 2017;130:722-31.
 36. Seiler M, Yoshimi A, Darman R, Chan B, Keaney G, Thomas M, *et al.* H3B-8800, an orally available small-molecule splicing modulator, induces lethality in spliceosome-mutant cancers. *Nat Med* 2018;24:497-504.
 37. Heuser M, Freeman SD, Ossenkoppele GJ, Buccisano F, Hourigan CS, Ngai LL, *et al.* 2021 update on MRD in acute myeloid leukemia: A consensus document from the European LeukemiaNet MRD working party. *Blood* 2021;138:2753-67.
 38. Tsai CH, Tang JL, Tien FM, Kuo YY, Wu DC, Lin CC, *et al.* Clinical implications of sequential MRD monitoring by NGS at 2 time points after chemotherapy in patients with AML. *Blood Adv* 2021;5:2456-66.
 39. Hou HA, Lin CC, Chou WC, Liu CY, Chen CY, Tang JL, *et al.* Integration of cytogenetic and molecular alterations in risk stratification of 318 patients with *de novo* non-M3 acute myeloid leukemia. *Leukemia* 2014;28:50-8.
 40. Chou SC, Tang JL, Hou HA, Chou WC, Hu FC, Chen CY, *et al.* Prognostic implication of gene mutations on overall survival in the adult acute myeloid leukemia patients receiving or not receiving allogeneic hematopoietic stem cell transplantations. *Leuk Res* 2014;38:1278-84.
 41. Platzbecker U, Middeke JM, Sockel K, Herbst R, Wolf D, Baldus CD, *et al.* Measurable residual disease-guided treatment with azacitidine to prevent haematological relapse in patients with myelodysplastic syndrome and acute myeloid leukaemia (RELAZA2): An open-label, multicentre, phase 2 trial. *Lancet Oncol* 2018;19:1668-79.
 42. Tiong IS, Hiwase DK, Abro E, Bajel A, Palfreyman E, Beligaswatte A, *et al.* Targeting molecular measurable residual disease and low-blast relapse in AML with venetoclax and low-dose cytarabine: A prospective phase II study (VALDAC). *J Clin Oncol* 2024;42:2161-73.
 43. Bhansali RS, Pratz KW, Lai C. Recent advances in targeted therapies in acute myeloid leukemia. *J Hematol Oncol* 2023;16:29.
 44. Montesinos P, Recher C, Vives S, Zarzycka E, Wang J, Bertani G, *et al.* Ivosidenib and azacitidine in IDH1-mutated acute myeloid leukemia. *N Engl J Med* 2022;386:1519-31.